Mechanism of Amyloid Removal in Patients With Alzheimer Disease Treated With Gantenerumab

Susanne Ostrowitzki, MD; Dennis Deptula, PhD; Lennart Thurfjell, PhD; Frederik Barkhof, MD; Bernd Bohrmann, PhD; David J. Brooks, MD, DSc; William E. Klunk, MD; Elizabeth Ashford, BSc; Kisook Yoo, PhD; Zhi-Xin Xu, MD; Hansruedi Loetscher, PhD; Luca Santarelli, MD

Background: Gantenerumab is a fully human anti-Aβ monoclonal antibody in clinical development for the treatment of Alzheimer disease (AD).

Objectives: To investigate whether treatment with gantenerumab leads to a measurable reduction in the level of Aβ amyloid in the brain and to elucidate the mechanism of amyloid reduction.

Design: A multicenter, randomized, double-blind, placebo-controlled, ascending-dose positron emission tomographic study. Additionally, ex vivo studies of human brain slices from an independent sample of patients who had AD were performed.

Setting: Three university medical centers.

Patients: Patients with mild-to-moderate AD.

Intervention: Two consecutive cohorts of patients received 2 to 7 infusions of intravenous gantenerumab (60 or 200 mg) or placebo every 4 weeks. Brain slices from patients who had AD were coincubated with gantenerumab at increasing concentrations and with human microglial cells.


Results: Sixteen patients with end-of-treatment positron emission tomographic scans were included in the analysis. The mean (95% CI) percent change from baseline difference relative to placebo (n=4) in cortical brain amyloid level was −15.6% (95% CI, −42.7 to 11.6) for the 60-mg group (n=6) and −35.7% (95% CI, −63.5 to −7.9) for the 200-mg group (n=6). Two patients in the 200-mg group showed transient and focal areas of inflammation or vasogenic edema on magnetic resonance imaging scans at sites with the highest level of amyloid reduction. Gantenerumab induced phagocytosis of human amyloid in a dose-dependent manner ex vivo.

Conclusion: Gantenerumab treatment resulted in a dose-dependent reduction in brain amyloid level, possibly through an effector cell–mediated mechanism of action.

Arch Neurol. Published online October 10, 2011. doi:10.1001/archneurol.2011.1538

In humans, amyloid plaque clearance by antiamyloid treatment was first suggested in autopsy cases following vaccination with Aβ42 (AN-1792; Elan Pharmaceuticals, Monksland, Athlone, County Westmeath, Ireland).1 However, because 6% of the study population developed meningoencephalitis,2 efficacy remained untested. A recent study using carbon 11–labeled Pittsburgh Compound B ([11C]PiB) positron emission tomography (PET) has shown that passive immunization can reduce the level of brain amyloid in vivo after 18 months of treatment,3 and this approach may be less prone to inducing severe neuroinflammation.4 The exact mechanism underlying amyloid reduction by immunotherapy has remained elusive.

We previously reported the development of gantenerumab, a potent and fully human anti-Aβ antibody that binds specifically to Aβ plaques.5 Gantenerumab has been studied in single- and multiple-dose phase 1 clinical trials (F. Hoffmann–La Roche Ltd, data on file). In the present study of patients with mild to moderate AD, we investigated the effect of up to 7 infusions of intravenous gantenerumab (60 or 200 mg) or placebo every 4 weeks on the level...
of brain Aβ amyloid as measured by [11C]PiB PET. Additionally, we report local effects of gantenerumab on brain magnetic resonance imaging (MRI) and provide an integrated analysis of results from the 2 imaging modalities. Furthermore, we link imaging results to data from an ex vivo assay in brain slices, all in an effort to elucidate the mechanism by which gantenerumab reduces the level of brain amyloid.

METHODS

PATIENTS

Data reported here are from a PET substudy of a multiple ascending dose (MAD) trial with gantenerumab. The clinicaltrials.gov identifier for the MAD study is NCT00531804. Complete methods and results from the MAD study will be reported separately; only select data related to the PET data are included here. To be eligible for the PET substudy, patients had to fulfill all entry criteria of the MAD study, with the following key criteria: 50 to 90 years of age, probable AD according to the National Institute of Neurological and Communicative Disorders and Stroke–Alzheimer’s Disease and Related Disorders Association criteria, a Mini-Mental State Examination score between 16 and 26 (inclusive), an MRI scan consistent with AD, and a modified Hachinski ischemia score of 4 or less. Stable symptomatic treatment of AD was allowed. Eligibility criteria specific to the PET substudy excluded patients who had been exposed to radiation in the past year or planned such exposure. Patients signed a written informed consent (cosigned by the patient’s next of kin or caregiver, if required by local regulations) prior to screening. The PET substudy was reviewed and approved by an independent ethics committee at each site as well as by the respective health authorities.

GANTENERUMAB

Gantenerumab is a human IgG1 with a high affinity for fibrillar Aβ. The original clone was derived from the MorphoSys HuCAL-Fab1 phage display Human Combinatorial Antibody Library ( Martinsried, Germany) and optimized by in vitro affinity maturation. Specificity for human Aβ present in senile plaques was demonstrated ex vivo by immunohistochemical staining of human brain sections at low picomolar concentrations. In vivo, gantenerumab crosses the blood-brain barrier and binds specifically and dose-dependently to Aβ plaques in PS2APP transgenic mice. Long-term treatment with gantenerumab over 5 months significantly decreased the amyloid plaque load in PS2APP mice assessed immunohistochemically.

RANDOMIZATION AND BLINDING

A subset of patients from 2 cohorts (a 60-mg cohort followed by a 200-mg cohort) of the MAD study participated in the PET substudy. Patients in each cohort were randomly assigned to receive either gantenerumab or placebo with a drug-to-placebo ratio of 4:1. Study site and sponsor personnel were blinded to treatment.

MAD STUDY

Patients were to receive up to 7 intravenous infusions of gantenerumab or placebo every 4 weeks. DNA samples were obtained for APOE genotyping. Magnetic resonance imaging monitoring included a 3-dimensional T1-weighted, T2*-weighted, and a fluid-attenuated inversion recovery (FLAIR) sequence. The instruments used for the clinical assessments included the Alzheimer’s Disease Assessment Scale–cognitive subscale, the Alzheimer’s Disease Assessment Scale–cognitive subscale, the Alzheimer’s Disease Assessment Scale–cognitive subscale, the Alzheimer’s Disease Assessment Scale–cognitive subscale, the Alzheimer’s Disease Assessment Scale–cognitive subscale, the Mini-Mental State Examination, a modified neuropsychological test battery, disability assessment for dementia, adverse event reporting, and laboratory tests.

PET SUBSTUDY

Positron emission tomographic imaging was performed at 3 sites using ECAT EXACT HR+ cameras (Siemens, Erlangen, Germany). Approximately 370 MBq of [11C]PiB (prepared as per local procedures) were administered as an intravenous bolus. The PET data were collected 60 to 90 minutes after the tracer injection. Frame-to-frame realignment was used to correct for any motion before a sum image was created. [11C]PiB summed images were coregistered to the patient’s baseline MRI scan, and MRI and PET data were spatially normalized into Montreal Neu-
The sample size was pragmatic rather than based on statistical power estimations. Approximately 35 patients from 4 dose cohorts were expected to participate. However, only 2 cohorts participated; a lower dose cohort (20 mg) was not included in the STATISTICAL ANALYSIS

The IN VITRO INTERACTION STUDY

To evaluate whether gantenerumab interferes with hydrogen 3–labeled \[^{[11C]}\text{PiB}\] binding to amyloid, an in vitro study was performed. Consecutive frozen sections from brains of 2 patients who had AD but were not in the clinical trial (Banner Sun Health Research Institute, University of Arizona, Sun City) were preincubated with up to 5000 ng/mL of gantenerumab to saturate antibody binding sites on amyloid plaques and were subsequently incubated with 1nM of \[^{[3H]}\text{PiB}\] to determine total bind-
ing. Consecutive sections were incubated with \[^{3}H\]-PiB in the presence of an excess unlabelled ligand (1µM) to determine non-specific binding. Images were assessed quantitatively using a Fujiﬁlm BAS-TR2025 phosphoimager (Tokyo, Japan).

EX VIVO PHAGOCYTOSIS ASSAY

To evaluate gantenerumab’s ability to clear amyloid plaques via phagocytosis, an ex vivo study was performed. Human primary microglia cells were freshly isolated from healthy human brain tissue obtained during tumor surgery (University Hospital Zurich, Switzerland). After gentle homogenization, microglia cells were enriched, removed from flasks, analyzed by a ﬂuorescence-activated cell sorter using anti-CD45, and used in the phagocytosis assay if more than 95% of the cells were positive for CD45.

Cortical brain tissue from patients who had AD but were not in the clinical trial (Braak stage VI, at Banner Sun Health Research Institute) was cryosectioned at a nominal thickness of 20 µm and placed onto culture dishes (Biocoat 40629; BD Biocoat, San Jose, California). Consecutive sections were preincubated with and without different concentrations of gantenerumab before the microglia cells were seeded at 1.5 × 10^6 cells/mL and cultured at 37°C for 3 days. After ﬁxation, Aβ plaques were detected by staining with an N-terminal speciﬁc mouse monoclonal antibody BAP-2 conjugated to Alexa Fluor 488 dye (Molecular Probes, Eugene, Oregon). An unrelated human IgG1 (PHP010; AbD Serotec, Raleigh, North Carolina) antibody served as a control.

Time-lapse live-cell imaging was done over 12 hours at an image frequency of every 10 minutes. Gantenerumab conjugated to Alexa Fluor 555 dye (Molecular Probes, Eugene, Oregon) was preincubated at 5 µg/mL, and microglia cells seeded, at conditions described above in a culture chamber attached to a Leica SP2 confocal microscope (Leica Microsystems, Buffalo Grove, Illinois).

RESULTS

PET STUDY

Reduction in Brain Amyloid Level After Treatment With Gantenerumab

In this PET study, 18 patients were randomly assigned to receive either placebo or gantenerumab (60 or 200 mg intravenously) (Figure 1 and Table 1). Owing to the early termination of dosing in the 200-mg cohort, not all patients received all 7 infusions (Table 2). Although the mean MMSE score was similar across groups at baseline, patients in the placebo group were younger and had lower brain Aβ amyloid levels (Tables 1 and 2). Hence, a statistical evaluation of the data was adjusted for baseline SUVR.

Table 2 and Figure 2 summarize the cortical composite SUVR at the end of treatment and the means of individuals’ changes from baseline. The actual mean (SD) changes were 0.24 (0.15) for the placebo group, 0.03 (0.24) for the 60-mg group, and −0.27 (0.45) for the 200-mg group. The mean (SD) percent change from baseline over total PiB signal was 11.0% (7.6%) for the placebo group, 2.1% (10.3%) for the 60-mg group, and −9.4% (14.0%) for the 200-mg group. The mean (SD) percent change from baseline over the specific PiB signal was 20.9% (15.6%) for the placebo group, 5.3% (19.7%) for
Figure 3. Effect of gantenerumab on amyloid load as indexed by standard uptake value ratios (SUVRs) using carbon 11-labeled Pittsburgh Compound B ([11C]PiB) positron emission tomography. Scatterplot shows percent change from baseline (specific [11C]PiB signal) in cortical composite SUVR over gantenerumab doses for all patients with an end-of-treatment scan who received gantenerumab (60 or 200 mg) or placebo every 4 weeks. The dose-response relationship is indicated by the linear regression line (% change in amyloid = 12.81 - 0.13 × dose) of the baseline-adjusted percent change residual value (vertical axis) vs actual dose of gantenerumab (horizontal axis).

Figure 4. Effect of gantenerumab on amyloid load as indexed by standard uptake value ratios (SUVRs) using carbon 11-labeled Pittsburgh Compound B ([11C]PiB) positron emission tomography. The median SUVR percent changes from baseline (specific [11C]PiB signal) by brain region are shown for patients who received infusions of intravenous gantenerumab (60 or 200 mg) or placebo every 4 weeks.

Figure 5. Magnetic resonance imaging (MRI) scans from an APOE ε4 homozygous patient. Images shown represent scans at baseline (A), during treatment (B), and after treatment (C) that were acquired using a fluid-attenuated inversion recovery sequence. The new area of hyperintensity on the scan performed during treatment (B) is most prominent in the right temporal lobe (arrow) and is consistent with inflammation or vasogenic edema. It first appeared on the scheduled MRI scan 2 weeks after the second drug infusion, was progressive for 6 weeks, and subsequently spontaneously completely resolved by week 17 (C).
the 60-mg group, and −14.9% (20.3%) for the 200-mg group. The observed mean (95% CI) treatment differences from the placebo group in percent change over the specific PiB signal were −15.6% (95% CI, −42.7% to 11.6%) for the 60-mg group and −35.7% (95% CI, −63.5% to −7.9%) for the 200-mg group. Adjusting for baseline SUVR, we found that a nonparametric analysis of covariance on this percent change suggested that the 200-mg group differed from the placebo group (P=.06). The dose dependency of the amyloid-reducing effect was indicated by the nonparametric linear regression analysis on the baseline-adjusted percent change values over the specific PiB signal: slope of −0.13 (r²=0.29; P=.03) (Figure 3).

Figure 6. Integrated analysis of amyloid positron emission tomography (PET) and magnetic resonance imaging (MRI). The PET and MRI scans from the 2 patients (ie, patients A and B) are shown. The baseline standard uptake value ratio (SUVR) images are superimposed on the baseline MRI scans (A and C), and the binary masks of the MRI (fluid-attenuated inversion recovery) findings as outlined by the expert reader are superimposed on the baseline structural MRI scans (B and D). The end-of-treatment SUVR maps are superimposed on the baseline MRI scans (E and G), and the difference maps of SUVRs at the end of treatment minus baseline are superimposed on baseline MRI scans (F and H). The late follow-up SUVR maps are superimposed on the baseline MRI scans (I and K), and the difference maps of SUVRs at late follow-up minus baseline are superimposed on the baseline MRI scans (J and L). Crosshairs indicate positioning of the MRI finding.
Changes were consistent across regions (Figure 4), except in the pons, which is a brain area known to have very limited amyloid deposition.9 Changes in subcortical white matter may indicate some gray matter contamination of this volume of interest. Although dose-dependent reductions in the level of amyloid were observed, no consistent treatment effects on cognitive measures were noted in this small group of patients treated for a short period of time. Moreover, individual changes in cognitive measures did not correlate with changes in levels of amyloid.

Greatest Reduction in Level of Amyloid in Areas of MRI-Detected Abnormality

Focal MRI signal changes were observed in 2 APOE ε4 homozygous patients following 2 and 4 doses of 200 mg of gantenerumab, respectively. Findings were most conspicuous on the FLAIR sequence and consistent with inflammation or vasogenic edema (Figure 5). They resolved spontaneously after discontinuation of dosing. Both patients also developed microhemorrhages (images not shown), and one of them was mildly symptomatic (headache, dizziness, gait instability, and tremor). All other patients (these include those in the MAD study) with such MRI changes were asymptomatic. Areas of high signal on FLAIR were often colocalized with prominent decreases in the SUVR (Figure 6). Patient A showed no overall reduction in the SUVR following 4 doses of gantenerumab (200 mg); however, a localized area of decreased SUVR in the area of the FLAIR signal in the right temporal lobe is shown in Figure 6E and F. This localized area of amyloid reduction was still present in the posttreatment PET scan acquired 6 months after complete resolution of the MRI finding (Figure 6I and J). Patient B (who happened to have the largest decrease in SUVR) showed an overall reduction in SUVR following 2 doses of gantenerumab (200 mg) with a unilateral amyloid reduction in the left caudate nucleus (Figure 6G and H), an area of focal high-FLAIR signal. This effect appeared essentially unchanged in the posttreatment PET scan performed 8 months after the MRI finding had completely resolved (Figure 6K and L). In both patients, amyloid reduction was greater in areas of FLAIR signal compared with the prespecified cortical composite volume of interest (Figure 7).

IN VITRO INTERACTION ASSAY

In vitro studies demonstrated that there was a lack of interference between [3H]-PiB and gantenerumab, and therefore these studies support the notion that in vivo changes in the SUVR that are based on [11C]PiB binding truly reflect changes in fibrillar plaque amyloid load.

EX VIVO PHAGOCYTOSIS ASSAY

A decrease in Aβ amyloid plaque in sections of brain that were incubated with microglia cells was dependent on the concentration of gantenerumab, with a slight effect at 50 ng/mL of gantenerumab and substantial plaque clearance at 500 and 5000 ng/mL (Figure 8). Live-cell imaging showed that a removal of fluorescent-labeled gantenerumab bound to amyloid deposits occurred within hours through active intracellular uptake by migrating microglia adjacent to amyloid plaques (Figure 8; videos, http://www.archneurol.com).
similar MRI findings in 2 patients (both carriers of APOE ε4/ε4) treated with 200 mg of gantenerumab. Although MRI cannot determine with certainty the underlying pathophysiology, this focal high-FLAIR signal was frequently colocalized with areas of higher amyloid reduction. Such local effects on amyloid PET could not be attributed to poor tracer penetration due to acute, local edema because they were still apparent 6 to 8 months after complete resolution of the MRI finding.

Several mechanisms for brain amyloid reduction by antiamyloid antibodies have been suggested. They include effector cell–mediated phagocytosis and direct dissolution of amyloid.16 Our observation of more prominent amyloid reduction in areas of increased FLAIR signal may provide clues as to the mechanism by which gantenerumab clears amyloid: (1) Microglial cells contain very low levels of Aβ in untreated patients with AD,17 whereas postmortem studies following treatment with AN-1792 suggest that antiamyloid antibodies lead to an increase in Aβ phagocytosis.17,18 Furthermore, results from the ex vivo assay reported herein support the hypothesis that gantenerumab clears amyloid plaques via Fc re-

Figure 8. Microglial phagocytosis of human amyloid plaques. Amyloid plaque staining of human Alzheimer disease (AD) brain sections in the absence of (A; scale bar, 100 µm) and after preincubation with gantenerumab followed by incubation with primary human microglia as effector cells (B-D; scale bars, 100 µm). Aβ amyloid plaques were decreased in the presence of human microglia after preincubation with gantenerumab in a concentration-dependent manner, with slight clearance of small plaques seen at 50 ng/mL (B) and substantial decrease of plaques at 500 ng/mL (C) and 5000 ng/mL (D). Live-cell imaging showed the removal of Alexa Fluor 555–conjugated gantenerumab (E and H; scale bars, 20 µm) by a migrating microglia cell adjacent to amyloid deposits depicted at start (E-G; scale bars, 20 µm) and 12 hours (H-J; scale bars, 20 µm). An example of a removed part of gantenerumab-stained amyloid is indicated by an arrowhead, and an example of newly phagocytosed gantenerumab within the migrating microglia cell is indicated by an arrow. Differential interference contrast (F and I) and merged (G and J) images are shown to follow the movement of the microglia cell (small arrowheads) and the intracellular uptake of gantenerumab at amyloid deposits over the incubation period.
ceptor/microglia-mediated phagocytosis, followed by lysosomal degradation as demonstrated for differentiated human macrophages. The colocalization of the focal FLAIR signal and amyloid reduction may be due to an exaggerated microglial response resulting in locally perturbed vascular permeability. (2) Direct dissolution of aggregated Aβ and subsequent Aβ drainage along the perivascular pathways may result in a transient increase in cerebral amyloid angiopathy. Accordingly, patients who received active Aβ immunization treatment were reported to have a significant increase in the level of Aβ42 (and Aβ40 to a lesser extent) in cerebral vessel walls at autopsy. When plaques are dissolved rapidly, clearance mechanisms may get saturated with a possible result of vasogenic edema. Also, in this instance, one might expect the MRI finding to more likely occur in or adjacent to areas with greater amyloid clearance, and both mechanisms may result in microhemorrhages.

In summary, although both clearance mechanisms may occur in parallel, the ex vivo data reported herein implicate phagocytosis as a more likely mechanism of amyloid reduction by treatment with gantenerumab. The FLAIR hyperintensities may be seen as instances of excessive pharmacological activity due to a high dose or more susceptible individuals (eg, carriers of the APOE ε4 genotype). Indeed, a lesser degree of Aβ amyloid reduction relative to placebo was observed in other brain areas and with a lower dose of gantenerumab in the absence of detectable FLAIR hyperintensities. This suggests that gantenerumab-induced amyloid lowering can be achieved without significantly perturbing vascular permeability through inflammation or blockage of Aβ clearance pathways when appropriate dosing is selected.

The main limitations of the present study are its small size and the unequal distribution of amyloid load at baseline between the treatment and placebo groups. Although statistical analysis methods were chosen to address these limitations, any conclusions are provisional in nature. Additionally, it is still unclear whether any reduction in brain amyloid level will translate into clinical efficacy. A phase 2 clinical trial is under way to investigate whether a clinical benefit can be achieved in gantenerumab-treated patients with prodromal AD.

Accepted for Publication: August 16, 2011. Published Online: October 10, 2011. doi:10.1001/archneurol.2011.1538

Correspondence: Luca Santarelli, MD, F. Hoffmann–La Roche Ltd, Neuroscience, Grenzacherstrasse 124, CH-4070 Basel, Switzerland (luca.santarelli@roche.com).

Author Contributions: Study concept and design: Ostrowitzki, Deptula, Bohrmann, Brooks, Ashford, Xu, and Santarelli. Acquisition of data: Deptula, Thurfjell, Barkhov, Bohrmann, and Xu. Analysis and interpretation of data: Ostrowitzki, Deptula, Thurfjell, Barkhov, Bohrmann, Brooks, Klunk, Yoo, Loetscher, and Santarelli. Drafting of the manuscript: Ostrowitzki, Deptula, Thurfjell, Bohrmann, Ashford, Yoo, and Santarelli. Critical revision of the manuscript for important intellectual content: Ostrowitzki, Deptula, Barkhov, Brooks, Klunk, Xu, Loetscher, and Santarelli. Statistical analysis: Yoo. Obtained funding: Santarelli. Administrative, technical, and material support: Ostrowitzki, Deptula, Bohrmann, Ashford, Loetscher, and Santarelli. Study supervision: Ostrowitzki, Deptula, Brooks, and Ashford.

Financial Disclosure: Drs Ostrowitzki, Deptula, Bohrmann, Ashford, Yoo, Xu, Loetscher, and Santarelli are full-time employees of Roche/F. Hoffmann–La Roche Ltd, and they may additionally hold Roche stock/stock options. Dr Thurfjell is a full-time employee of GE Healthcare. Dr Barkhof is receiving consulting fees from pharmaceutical companies, including F. Hoffmann–La Roche Ltd. Dr Brooks holds a part-time position as a Senior Neurologist with GE Healthcare, which holds a license agreement with the University of Pittsburgh based on the [14C]PIB PET technology described in this study. Dr Klunk is the co-inventor of this technology and, as such, has a financial interest in this license agreement. GE Healthcare has provided grant support and consultant fees to Dr Klunk for studies unrelated to this work, and Dr Klunk has received consultant fees/honoraria related to this study from Roche. During the past 12 months, Dr Brooks has received consultancy fees from Synosia, Schering Plough, Amsterdam Molecular Therapeutics, Biogen Idec, NeuroNova, Shire, Genentech, and Janssen and honoraria from Teva, UCB, GlaxoSmithKline, and Orion Pharma.

Funding/Support: This work was supported by F. Hoffmann–La Roche Ltd.

Online-Only Material: The eTable and videos are available at http://www.archneurol.com.

Additional Contributions: We thank the following investigators for supporting this PET study: Niels Andreasen, MD (Karolinska Institutet, Huddinge University Hospital, Stockholm, Sweden), Roger Bullock, MD (Kingshill Research Centre, Swindon, England), Mark Dale, MD (Memory Assessment Centre, Blackpool, England), Adriana Lammertsma, PhD (Nuclear Medicine and PET Research, VU University Medical Center, Amsterdam, Netherlands), and Philip Scheltens, MD (Alzheimer Centre, VU University Medical Center, Amsterdam, Netherlands). We also thank the following Roche employees: Sumu Sethi, PhD, for her role in monitoring the study conduct, and Françoise Gerber, Krisztina Oroszlan, and Juerg Messer for their support in brain sectioning, in performing immunohistochemical stainings, confocal microscopy, and autoradiography. We thank Ursula Puentener, PhD, for isolating and enriching primary human microglia cells. We especially thank the patients and their caregivers for their support in this study.

REFERENCES


